

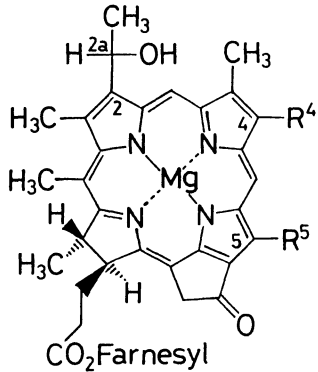
FT-IR Studies on the Aggregation of Bacteriochlorophyll c from Chlorobium limicolaKaku UEHARA,\* Yukihiro OZAKI,<sup>†</sup> Kouichi OKADA,<sup>†</sup> and John M. OLSON<sup>††</sup>Department of Applied Chemistry, College of Engineering,  
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Aggregates of three homologs of bacteriochlorophyll c from Chlorobium limicola in water-saturated carbon tetrachloride and in solid films were studied by means of FT-IR. Four keto-carbonyl stretching peaks were observed at 1703, 1682-4, 1649-51, and 1641-3  $\text{cm}^{-1}$  corresponding to free C=O (monomer), free C=O (dimer, tetramer), coordinated C=O (dimer, tetramer), and coordinated C=O (polymer), respectively.

Bacteriochlorophyll c (BChl c) in Chlorobium limicola (green sulfur bacterium) has several homologs including the 2-(R)-(1-hydroxyethyl)-4-ethyl-5-ethyl-homolog (EEF-BChl c), the 2-(R)-(1-hydroxyethyl)-4-n-propyl-5-ethyl-homolog (PEF-BChl c), and the 2-(S)-(1-hydroxyethyl)-4-isobutyl-5-methyl/ethyl-homolog (iBM/EF-BChl c).<sup>1)</sup> Table 1 shows the structural assignments for BChl c. These BChl c homologs occur in the chlorosome as higher aggregates and act as the light harvesting antenna.<sup>2)</sup> Olson and Pedersen<sup>3)</sup> reported that PEF-BChl c (R-configuration) forms dimers absorbing ca. 680 nm and tetramers (Olson and Cox, unpublished) absorbing at ca. 705-710 nm in  $\text{CH}_2\text{Cl}_2$ ,  $\text{CHCl}_3$  and  $\text{CCl}_4$ . On the other hand, iBM/EF-BChl (S-configuration) in  $\text{CCl}_4$  forms aggregates whose absorption (747 nm maximum) and circular dichroism spectra resemble those of the chlorosome. Uehara and Olson (unpublished) identified the forms of BChl c in water-saturated carbon tetrachloride ( $\text{H}_2\text{O}$ -satd  $\text{CCl}_4$ ) as monomer, dimer, tetramer, and polymer from the analysis of visible absorption spectra by the Simplex curve-fitting method (Olson and Cox, unpublished). Ballschmiter and Katz<sup>4,5)</sup> have previously reported the infrared spectra of dimers, oligomers, and water-coordinated aggregates (polymers) of various chlorophylls in solution and in solid films.

In the present paper, we report a new finding that PEF- and iBM/EF-BChl c homologs form polymers in solid films from  $\text{H}_2\text{O}$ -satd  $\text{CCl}_4$  solution. Fourier transform-infrared (FT-IR) spectra of  $\text{H}_2\text{O}$ -satd  $\text{CCl}_4$  solutions of EEF-, PEF-, and iBM/EF-BChl c homologs are also presented.

Chlorobium limicola f. thiosulfatophilum 6230 (Tassajara) was grown, and BChl c

Table 1. Structural assignments for the BChl c


The chemical structure of BChl c is shown, featuring a central magnesium atom coordinated to four nitrogen atoms in a porphyrin-like ring. The structure includes methyl groups at positions 2, 3, 4, and 5, a hydroxyl group at C-2a, and a farnesyl ester group at C-6. Substituents R<sup>4</sup> and R<sup>5</sup> are attached to the ring at positions 4 and 5, respectively.

BChl <u>c</u>	R <sup>4</sup>	R <sup>5</sup>	Configuration at C-2a
EEF-	Et	Et	R
PEF-	n-Pr	Et	R
iBM/EF-	i-Bu	Me/Et	S

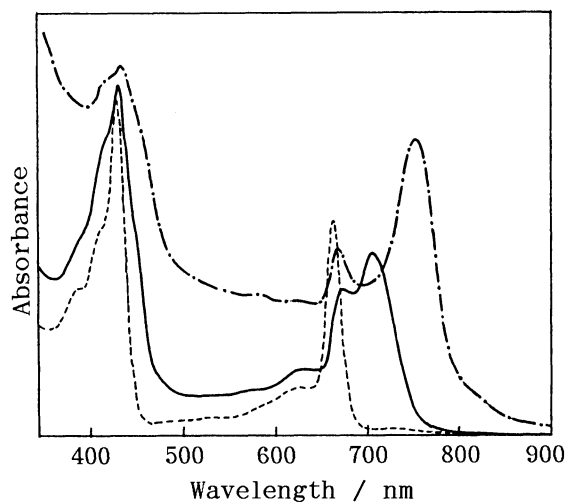


Fig. 1. Absorption spectra of PEF-BChl c in the H<sub>2</sub>O-satd CCl<sub>4</sub> (solid), in solid film on CaF<sub>2</sub> from the solution (dot-dash) and in the H<sub>2</sub>O-satd CCl<sub>4</sub> solution after addition of 1:200 methanol (dash).

was extracted from wet cells as described previously.<sup>3,6)</sup> FT-IR and absorption spectra were recorded with a JEOL JIR-100 FT-IR spectrometer and a Shimadzu UV-360 spectrophotometer, respectively. H<sub>2</sub>O-satd CCl<sub>4</sub> was used as the solvent for liquid film samples (0.995 mM, 1.14 mM, and 1.45 mM for iBM/EF-, EEF-, and PEF-BChl c, respectively). A Spectra Tech Contact Sampler was employed for horizontal FT-IR-ATR measurement (45°, ZnSe) of solid BChl c films from the H<sub>2</sub>O-satd CCl<sub>4</sub> solution. A CaF<sub>2</sub> plate was used for visible absorption spectrum measurements of solid BChl c films.

Figure 1 shows absorption spectra of PEF-BChl c in H<sub>2</sub>O-satd CCl<sub>4</sub> (108 μM) and of the solid film from solution on CaF<sub>2</sub>. In the H<sub>2</sub>O-satd CCl<sub>4</sub> the spectral pattern of the Q<sub>y</sub> band with two absorption maxima at 673 and 705 nm is similar to that for PEF-BChl c in dry CCl<sub>4</sub> which can be deconvoluted into three components; monomer (662 nm; see Fig. 1, dash line), dimer (680 nm), and tetramer (705 nm)<sup>3)</sup> (Olson and Cox, unpublished). On the other hand, the absorption spectrum of the solid film shows a large polymeric band at 750 nm with a small monomeric band at 667 nm. A marked red-shift of the Q<sub>y</sub> band from 705 nm to 750 nm upon going from solution to solid film indicates the transformation of dimer and tetramer of PEF-BChl c into polymer.

As shown in Fig. 2 and Table 2, four keto C=O stretching peaks were observed at 1703, 1682-4, 1649-51, and 1641-3 cm<sup>-1</sup>. The bands at 1703 and 1682-4 cm<sup>-1</sup> can be assigned to the free keto C=O.<sup>4,7)</sup> A marked low-wavenumber shift of the keto C=O peak to lower wavenumber arises from the coordination of the keto C=O to the Mg atom of another BChl c molecule or from the hydrogen bonding with the hydroxy group of another BChl c or water molecule involved in the aggregation<sup>6,8)</sup> to form dimer, tetramer, or polymer. The bands at 1649-51 and 1641-3 cm<sup>-1</sup> might be assigned to coordi-

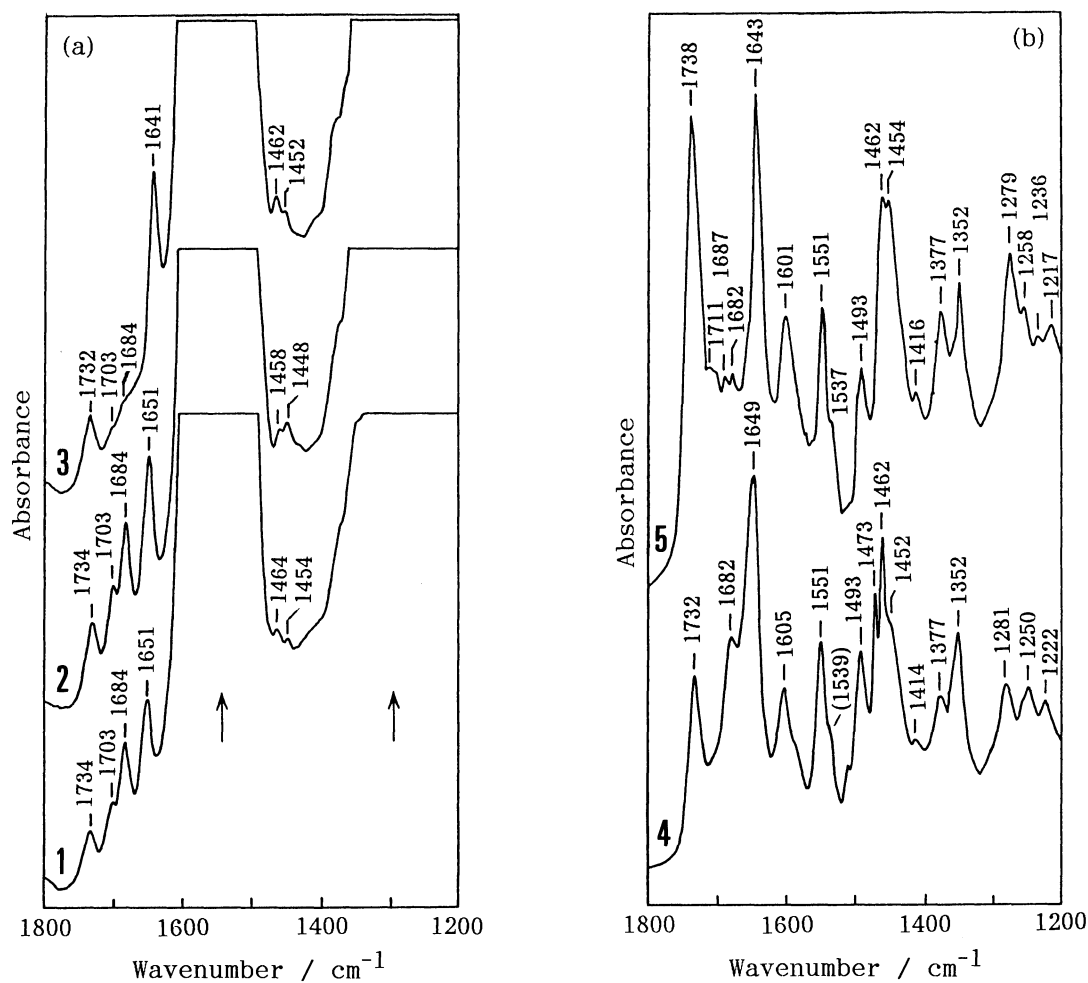


Fig. 2. FT-IR spectra of BChl c homologs in H<sub>2</sub>O-satd CCl<sub>4</sub> (a: 1, EEF-; 2, PEF-; 3, iBM/EF-) and in solid films on ZnSe from the solutions (b: 4, EEF-; 5, PEF-). Arrow: absorption band of CCl<sub>4</sub>.

Table 2. Comparison of carbonyl peak wavenumbers (cm<sup>-1</sup>) in FT-IR spectra of BChl c in H<sub>2</sub>O-satd CCl<sub>4</sub> and in solid films from solution on ZnSe

Tentative assignment	in H <sub>2</sub> O-satd CCl <sub>4</sub> solution			in solid film		
	EEF-	PEF-	iBM/EF-	EEF-	PEF-	iBM/EF-
ester C=O	1734 s <sup>e)</sup>	1734 s	1732 s	1732 s	1738 s	1734 s
free keto C=O <sup>a)</sup>	1703 m	1703 m <sup>f)</sup>	1703 sh <sup>g)</sup>	-	-	-
free keto C=O <sup>b)</sup>	1684 s	1684 s	1684 sh	1682 m	1682 w <sup>h)</sup>	-
coordinated keto C=O <sup>c)</sup>	1651 s	1651 s	-	1649 s	-	-
coordinated keto C=O <sup>d)</sup>	-	-	1641 s	-	1643 s	1641 s

a)(monomer). b)(1/2 from dimer + 1/4 from tetramer). c)(1/2 from dimer + 3/4 from tetramer). d)(polymer). e)s, strong. f)m, medium. g)sh, shoulder. h)w, weak.

nated keto C=O of oligomer<sup>4)</sup> (dimer, tetramer) and polymer (hydrated aggregates)<sup>5)</sup>, respectively. From the dimer-tetramer ratio in EEF- and PEF-BChl c solutions, the band at 1682-4 cm<sup>-1</sup> must contain contributions from both dimer and tetramer (1/2 from dimer + 1/4 from tetramer) and the 1649-51 cm<sup>-1</sup> band must also contain contributions from both dimer and tetramer (1/2 from dimer + 3/4 from tetramer). On the other hand, the IBM/EF-homolog showed a strong polymeric peak at 1641 cm<sup>-1</sup> with very weak free keto C=O peaks at 1703 and 1684 cm<sup>-1</sup>. In solid films, however, both the IBM/EF- and the PEF-BChl c homolog showed a strong polymer peak at 1641 and 1643 cm<sup>-1</sup>, respectively. It is important to note that resonance Raman spectrum of BChl c in the Chlorobium limicola chlorosome shows a single, sharp band at 1639-1640 cm<sup>-1</sup>.<sup>8)</sup> The ester C=O stretching peak was observed at 1732-8 cm<sup>-1</sup> for all three BChl c homologs in solution or solid film, suggesting almost no interaction of the ester C=O with a Mg atom or OH-group of another BChl c molecule. The presence of bands for C=C stretching vibrations at 1551 cm<sup>-1</sup> (s, strong), 1537-9 cm<sup>-1</sup> (sh, shoulder) and 1601-5 cm<sup>-1</sup> (s) do not fall within the 6-coordination ranges and suggest 5-coordination of the Mg atom.<sup>8-10)</sup> The asymmetric 6-coordinate model<sup>11)</sup> probably can be excluded. The antiparallel-chain model with 5-coordinate Mg but involving both the 2-hydroxyethyl and 9-keto groups<sup>12)</sup> is preferred. More detailed studies on the formation and structure of polymeric BChl c aggregates are now in progress.

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#### References

- 1) K. M. Smith, G. W. Craig, L. A. Kehres, and N. Pfennig, *J. Chromatogr.*, **281**, 209 (1983).
- 2) K. M. Smith and L. A. Kehres, *J. Am. Chem. Soc.*, **105**, 1387 (1983).
- 3) J. M. Olson and J. P. Pedersen, *Photosynth. Res.*, **25**, 25 (1990).
- 4) K. Ballschmiter and J. J. Katz, *J. Am. Chem. Soc.*, **91**, 2661 (1969).
- 5) K. Ballschmiter and J. J. Katz, *Biochim. Biophys. Acta*, **256**, 307 (1972).
- 6) J. M. Olson, K. D. Philipson, and K. Sauer, *Biochim. Biophys. Acta*, **292**, 206 (1973).
- 7) Y. Koyama, Y. Umemoto, A. Akamatsu, K. Uehara, and M. Tanaka, *J. Mol. Struct.*, **146**, 273 (1986).
- 8) M. Lutz and G. van Brakel, "Green Photosynthetic Bacteria," ed by J. M. Olson et al., Plenum Press, New York (1988), p. 23.
- 9) M. Fujiwara and M. Tasumi, *J. Phys. Chem.*, **90**, 250, 5646 (1986).
- 10) T. Nozawa, T. Noguchi, and M. Tasumi, *J. Biochem.*, **108**, 737 (1990).
- 11) D. C. Brune, T. Nozawa, and R. E. Blankenship, *Biochemistry*, **26**, 8644 (1986).
- 12) M. Mimuro, T. Nozawa, N. Tamai, K. Shimada, I. Yamazaki, Su Lin, R. S. Knox, B. P. Wittmerhause, D. C. Brune, and R. E. Blankenship, *J. Phys. Chem.*, **93**, 7503 (1989).

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